Histopathological Effects of Fibrin Glue and Cyanoacrylate on the Maxillary Sinus

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Abstract

Objective: To compare the histopathological effects of fibrin glue (FbGl) and cyanoacrylate (CyAc) on the maxillary sinus mucosa.

Methods: Twenty rabbits were divided into two groups of 10, and surgical defects were created in the maxillary sinuses. The right maxillary sinus was treated with FbGl in one group and with CyAc in the other group. As a control, the left maxillary sinuses of all rabbits were treated with sterile saline solution. One rabbit treated with CyAc died during the study and was excluded. On postoperative day 21, all animals were sacrificed. Maxillary sinus mucosa samples were studied to determine the extent of inflammation and fibrosis, foreign body reaction, cilia loss, increased osteogenesis in bony structures under the mucosa, and loss of serous glands.

Results: The FbGl group differed significantly from the CyAc and control groups in terms of a high degree of inflammation (p<0.001), fibrosis (p<0.001), foreign body reaction (p<0.001), cilia loss (p<0.001), and serous gland loss (p<0.001). In terms of osteogenesis, there were no significant differences between the FbGl and CyAc groups (p=0.650), while there was a significant (p=0.002) difference between these two groups and the control group.

Conclusion: Histopathologically, CyAc had fewer side effects than FbGl. Further clinical studies are needed to demonstrate the validity of these results in humans.

Keywords: Animal experimentation, maxillary sinus, fibrin glue, cyanoacrylate, histopathology

Introduction

The human body can often repair soft and hard tissue wounds, and this can be facilitated by fastening the tissues mechanically with sutures or staples (1). In nasal and functional endoscopic sinus surgery (ESS), the limited surgical field and the need to use minimally invasive procedures make it difficult to use sutures (the gold standard), particularly in mucosal regions (2). Wound closure, skin grafting, and the fixation of transplants and implants are vital to ensure surgical success. Tissue glues are also used to achieve closure and are currently the method of choice in dural plasty, haemostasis, and skin adhesion (1). However, wound closure systems using adhesives in otorhinolaryngology require more research.

During functional ESS, iatrogenic wounds often develop in the lamina cribrosa, particularly in the anterior ethmoid region. The lamina cribrosa is fragile, and defects and dehiscence can develop from the area of attachment of the median concha to the skull base and in the anterior and posterior ethmoidal artery channels. If an iatrogenic injury leads to cerebrospinal fluid leakage, the fistula can be closed with muscle, bone wax, fascia, a bony plate, a mucoperichondrial flap, a dural flap, tissue glue, or fat tissue (2, 3).

It is important that the material used for closure be inexpensive, readily available, easy to use, and highly compatible with the tissue. Many authors have used fibrin glue (FbGl) or a cyanoacrylate (CyAc) to adhere different tissues in both clinical and experimental studies. Sakagami et al. (4) successfully used FbGl as an underlay after myringoplasty, Grayeli et al. (5) used it after facial nerve anastomosis, and Vries et al. (6) used it after surgery on nerves. However, a risk of viral transfer has been reported (7, 8). In addition, the glue is relatively expensive and a technician may be needed to prepare autologous FbGl before the operation (9).

Pineros-Fernandez et al. (10) used CyAc to repair peripheral nerves, Bruns et al. (11) used it to repair skin lacerations, and Gulalp et al. (12) used it to
repair incisions of the skin and mucosa. Several papers have reported satisfactory aesthetic and functional results using CyAc (10-12). Some studies have reported increased inflammation and fibrosis, but primarily, when first-generation short-chain materials were used. During the degradation of CyAc polymers, formaldehyde is released and the extent of CyAc histotoxicity is proportional to the amount of formaldehyde produced (13). The most commonly used CyAcS, ordered from good to bad in terms of tissue tolerance, are n-decyl-, n-octyl-, n-heptyl-, n-buty1-, isobutyl-, and methyl-cyanoacrylate (14). When applied to the skin, the strength of n-octyl-cyanoacrylate is thrice that of n-buty1-cyanoacrylate, and it is almost as strong as 5.0 monofilament nylon suture (15). Therefore, we used n-octyl-cyanoacrylate in this study.

No study has compared the histopathological effects of tissue adhesives on the paranasal sinus mucosa. Therefore, we compared the effects of FbGl glue and CyAc on the maxillary sinus mucosa of rabbits.

Methods
We used 20 14- to 16-week-old New Zealand rabbits weighing 2.5–3 kg. All were fed a standard laboratory diet. The rabbits were kept in an animal room and monitored for several days before surgery to confirm that they were in good health. The animals were divided into two groups of 10 to study the histopathological effects of FbGl and CyAc on the maxillary sinus. All rabbits were anaesthetised with intramuscular (IM) ketamine HCl (Ketalar 10 mg/mL, (Ketalar; Pfizer, İstanbul, Turkey)) and xylazine (5 mg/kg). Each surgical site was scrubbed with povidone iodine thrice. The periosteum was reached via a 2-cm vertical incision commencing at the midsection of the nasal dorsum. Then, the periosteum was elevated off the bone via a vertical incision. Using a 2-mm diameter drill, two holes (on the left and right) were drilled into the front wall of the maxillary sinus; the holes were offset by about 1 cm from the midsection of the nasal bone. In the FbGl group (n=10), the right maxillary sinus was completely filled with FbGl (Tisseel®, Baxter AG; Vienna, Austria), and in the CyAc group (n=9), it was filled with wicking-type octyl-2-cyanoacrylate (Super Glue™; Osaka, Japan). As a control, the left maxillary sinuses of all rabbits were filled with sterile saline solution (n=19). For the FbGl group, the thrombin and fibrinogen components of FbGl in separate injectors were mixed in equal amounts and applied immediately. Then, the defects were covered with the periosteum and skin. After surgery, each incision was closed withatraumatic 3/0 silk suturing. During the operation, each rabbit was given 5 mg/kg ceftriaxone IM. One rabbit treated with CyAc died during the study and was excluded. Twenty-one days after the operation, all animals were euthanized, and the periosteum was opened as described above. Each maxillary sinus was extracted in the sagittal plane using an electric saw and studied.

Histopathological Tissue Investigations
Both sides of each maxillary sinus were fixed in 10% (v/v) formaldehyde for 24 h, decalcified in an electrolytic device in 15% (v/v) formic acid for 20 days, paraffinized, cut into 5-µm-thick slices, stained with haematoxylin–eosin (H–E), and observed under a light microscope to determine the extent of inflammation, fibrosis, foreign body reaction, cilia loss, osteogenesis of bony structures under the mucosa, and loss of serous glands.

The extent of inflammation was graded on the basis of lymphocyte and macrophage infiltration, tissue damage, and repair. Fibrosis was evaluated on the basis of the migration of fibroblasts
into the wound, cell proliferation, and storage of the extracellular matrix. Histopathological evaluation included analysis of foreign body reactions caused by activated macrophages, cilia loss, type 1 collagen levels, and abnormal development (osteogenesis and serous gland loss).

Statistical Analysis

Statistical analysis were performed using SPSS 19.0 (SPSS; Chicago, IL, USA). Descriptive statistics are expressed as the frequency and percentage. The chi-square test or Fisher's exact test was used to determine the differences among the three groups. A p-value of less than 0.05 was considered statistically significant for all tests.

Ethics/Patient Consent

Our study was approved by the Animal Ethics Committee of Bülent Ecevit University, Zonguldak, Turkey. All animals were treated according to the Helsinki Universal Declaration of Animal Rights.

Results

Inflammation

The FbGl group had significantly more inflammation than the CyAc (p=0.009) and control (p<0.001) groups. Figure 1 shows the extent of medium inflammation in the FbGl group. There were no significant (p=0.234) differences in inflammation between the CyAc and control groups (Table 1, Figure 2).

Fibrosis

Table 2 summarizes the extent of fibrosis in the lamina propria. There was 100% fibrosis in the FbGl group, 33% in the CyAc group, and none in the control group. Figure 3 shows the fibrosis in the FbGl group. There was significantly (p<0.001) more fibrosis in the FbGl group and significantly (p<0.026) more fibrosis in the CyAc group than in the control group (Tables 2, 3).

Foreign body reaction

Only the FbGl group showed a foreign body reaction (Table 2, Figure 4).

Cilia Loss

Cilia loss occurred in 90% of the rabbits in the FbGl group and in 33% of the rabbits in the CyAc group (Table 2); the difference was statistically significant (p<0.001). The difference between the CyAc and control groups was also significant (p=0.026).

Osteogenesis

Both the FbGl (50%) and CyAc (33%) groups showed significantly (p=0.002) more osteogenesis than the control group (0%) (Table 2), although the difference between the FbGl and CyAc groups was not significant (p=0.650). Figure 3 shows osteogenesis in the FbGl group.

Serous Gland Loss

There was no loss of serous glands in the controls, while a significant (p<0.001) loss was apparent in the FbGl (100%) and CyAc (33%) groups (Table 2). The difference between the CyAc and control groups was not significant (p=0.084).

Discussion

We found that FbGl increased inflammation. Erkan et al. (16) used FbGl to attach a nasal mucoperichondrial flap to the septal cartilage of the nasal mucosa of albino Vienna rabbits and noted histologically apparent inflammation. In addition, the mucosal thickness increased, while the thicknesses of the perichondrium

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**Table 2. Fibrosis, foreign body reactions, ciliary loss, osteogenesis, and serous gland loss in all groups**

<table>
<thead>
<tr>
<th></th>
<th>FbGl group n=10</th>
<th>CyAc group n=9</th>
<th>Control group n=19</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fibrosis</td>
<td>positive 10</td>
<td>3 33.3%</td>
<td>0 0%</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>none 0</td>
<td>0 0%</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Foreign body reaction</td>
<td>positive 7</td>
<td>0 0%</td>
<td>0 0%</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>none 3</td>
<td>9 100%</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Ciliary loss</td>
<td>positive 9</td>
<td>3 33.3%</td>
<td>0 0%</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>none 1</td>
<td>6 66.6%</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Osteogenesis</td>
<td>positive 5</td>
<td>3 33.3%</td>
<td>0 0%</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>none 5</td>
<td>6 66.6%</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Loss of serous glands</td>
<td>positive 10</td>
<td>3 33.3%</td>
<td>1 5.3%</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>none 0</td>
<td>6 66.6%</td>
<td>18 94.7%</td>
<td></td>
</tr>
</tbody>
</table>

FbGl: fibrin glue; CyAc: cyanoacrylate

**Table 3. Statistical analysis of fibrosis, foreign body reactions, ciliary loss, osteogenesis, and serous gland loss between CyAn and control groups**

<table>
<thead>
<tr>
<th></th>
<th>CyAc group n=9</th>
<th>Control group n=19</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Degree of inflammation</td>
<td>7 (light)</td>
<td>18 (light)</td>
<td>77.8%</td>
</tr>
<tr>
<td></td>
<td>2 (medium)</td>
<td>1 (medium)</td>
<td>94.7%</td>
</tr>
<tr>
<td>Fibrosis</td>
<td>positive 3</td>
<td>0 0%</td>
<td>22.2%</td>
</tr>
<tr>
<td></td>
<td>none 6</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Foreign body reaction</td>
<td>positive 0</td>
<td>0 0%</td>
<td>22.2%</td>
</tr>
<tr>
<td></td>
<td>none 9</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Ciliary loss</td>
<td>positive 3</td>
<td>0 0%</td>
<td>22.2%</td>
</tr>
<tr>
<td></td>
<td>none 6</td>
<td>19 100%</td>
<td></td>
</tr>
<tr>
<td>Osteogenesis</td>
<td>positive 3</td>
<td>0 0%</td>
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<td></td>
<td>none 6</td>
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<td>Loss of serous glands</td>
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<td>18 94.7%</td>
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</tbody>
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FbGl: fibrin glue; CyAc: cyanoacrylate
cartilage decreased, cartilage damage was evident, ciliary and goblet cell numbers decreased, and fibrosis developed. Altuntas et al. (3) evaluated the histopathological effects of FbGl on the maxillary sinus mucosa of rats. After 21 days, no significant difference between FbGl and saline was noted in terms of the extent of inflammation. In contrast, compared with controls, we observed a significant (p<0.001) increase in inflammation when FbGl was applied.

We also observed an increase in inflammation (compared with controls) when CyAc was applied, but this was not significant (p=0.234). We could not find any study that applied CyAc to the respiratory mucosa of rabbits.

In a similar study, Alkan et al. (17) used n-butyryl-cyanoacrylate to fix the septum to the anterior nasal spine and found no significant foreign body reaction histopathologically in comparison with a control group. Egemen et al. (20) used FbGl and suturing to secure a vascular anastomosis and compared this with suturing only in rats. The foreign body reaction was reduced using the glue/suturing combination. In our study, we did not find any foreign body reaction when CyAc was used, while FbGl produced a significant (p<0.001) foreign body reaction attributable to the sensitivity of the nasal mucosa relative to that of the vascular wall.

In terms of the numbers of cilia and serous cells, the FbGl group showed significantly greater losses than the CyAc or control group. In the study of Erkan et al. (16), in which mucoperichondrial flaps were fixed to the mucosal septal cartilage in rabbits using FbGl, similar losses were apparent. Choi et al. (21) used CyAc to close sinus membrane perforations in rabbits and observed that the extent of the extracellular matrix but not serous gland numbers decreased. This was attributed to the regeneration of subepithelial serous glands. We also observed significant (p<0.001) losses of cilia and serous cells (only 33%) in CyAc compared with controls, and FbGl was associated with significant losses in both components (p<0.001) compared with CyAc. This may be explained by tissue degeneration in the former and tissue regeneration in the latter.

Kania et al. (22) observed that FbGl accelerated bone formation under osteogenic conditions in a rabbit femoral defect model. In our study, there was no significant (p=0.650) difference between the FbGl and CyAc groups but a significant (p=0.002) difference between both these groups and controls. Overall, the increase in osteogenesis was more noticeable in the FbGl group (50%).

FbGl and CyAc have been used to repair various types of tissues. Petter-Puchner et al. (23) found no differences in
terms of air penetration in the lung tissue of rabbits when FbGI and CyAc were used. However, FbGI was preferred because CyAc was associated with significantly higher levels of inflammation and elevated wound temperatures in the early stages of healing. Wieken et al. (24) compared FbGI and CyAc in the context of neural anastomosis in a rat model. CyAc caused inflammation, reminiscent of a foreign body reaction, and decrease in the neural diameter by 66%. Consequently, CyAc was not preferred for peripheral neural repair. In contrast, we found that CyAc was better than FbGI in all aspects studied. This may be attributable to the particular structural features of the mucosal tissue and the microvascular density therein.

This study had some weaknesses. The first was the side effects of maxillary sinus obliteration caused by FbGI and CyAc. The maxillary sinus mucosa and its aeration were preserved in the control group but not in the FbGI and CyAc groups. Another important weakness is that we did not evaluate the effects of FbGI and CyAc on the nasal mucosa.

Conclusion
Histopathologically, CyAc had fewer side effects than FbGI. FbGI is also relatively expensive. Before either agent can safely be used on the paranasal sinus mucosa, they must be evaluated in larger groups and the long-term histopathological effects on various tissues should be explored.

Ethics Committee Approval: Ethics committee approval was received for this study from the ethics committee of Animal Ethics Committee of Bilent Ecevit University.

Informed Consent: Not required in this study.

Peer-review: Externally peer-reviewed.


Conflict of Interest: No conflict of interest was declared by the authors.

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References
1. Schneider G. Tissue adhesives in otorhinolaryngology. Laryngorhinootologie 2009; 88: S156-64. [CrossRef]
